of starch TS). Perform a blank determination.

Each mL of 0.05 mol/L iodine VS = 1.5013 mg of  $CH_2O$ 

Containers and storage Containers—Tight containers. Storage—Light-resistant.

#### Formalin Water

ホルマリン水

Formalin Water contains not less than 0.9 w/v% and not more than 1.1 w/v% of formaldehyde (CH<sub>2</sub>O: 30.03).

#### Method of preparation

| Formalin                | 30 mL                 |
|-------------------------|-----------------------|
| Water or Purified Water | a sufficient quantity |
|                         | To make 1000 mL       |

Prepare by mixing the above ingredients.

**Description** Formalin Water is a clear, colorless liquid. It has a slight odor of formaldehyde.

It is almost neutral.

Assay Transfer 20 mL of Formalin Water, measured exactly, to a 100-mL volumetric flask containing 2.5 mL of 1 mol/L sodium hydroxide VS, and add water to make 100 mL. Pipet 10 mL of this solution, and proceed as directed in the Assay under Formalin.

Each mL of 0.05 mol/L iodine VS = 1.5013 mg of  $CH_2O$ 

Containers and storage Containers—Tight containers.

## Forsythia Fruit

Forsythiae Fructus

レンギョウ

Forsythia Fruit is the fruit of Forsythia suspensa Vahl or Forsythia viridissima Lindley (Oleaceae).

**Description** Ovoid to long ovoid capsule, 1.5-2.5 cm in length, 0.5-1 cm in width, with acute apex, and sometimes with a peduncle at the base; externally light gray to dark brown, scattered with light gray and small ridged dots, and with two longitudinal furrows; a capsule dehiscing along the longitudinal furrows has the apexes bent backward; the inner surface of dehisced pericarp is yellow-brown in color, with a longitudinal partition-wall in the middle; seeds, slender and oblong, 0.5-0.7 cm in length, and usually with a wing. Odor, slight; tasteless.

**Identification** (1) To 0.2 g of pulverized Forsythia Fruit add 2 mL of acetic anhydride, shake well, allow to stand for 2 minutes, and filter. To 1 mL of the filtrate add gently 0.5 mL of sulfuric acid to form two layers: a red-purple color develops at the zone of contact.

(2) To 1 g of pulverized Forsythia Fruit add 10 mL of

methanol, warm on a water bath for 2 minutes, and filter. To 5 mL of the filtrate add 0.1 g of magnesium in ribbon form and 1 mL of hydrochloric acid, and allow to stand: a light red to yellow-red color develops.

**Purity** (1) Branchlet—The amount of branchlets contained in Forsythia Fruit does not exceed 5.0%.

(2) Foreign matter—The amount of foreign matter other than branchlets contained in Forsythia Fruit does not exceed 1.0%.

Total ash Not more than 5.0%.

Extract content Dilute ethanol-soluble extract: not less than 10.0%.

### Gambir

Gambir

アセンヤク

Gambir is the dried aqueous extract prepared from the leaves and young twigs of *Uncaria gambir* Roxburgh (*Rubiaceae*).

**Description** Brown to dark brown, brittle mass; inside light brown. Odor, slight; taste, extremely astringent and bitter.

**Identification** (1) To 0.2 g of pulverized Gambir add 10 mL of water, warm in a water bath for 5 minutes with occasional shaking, and filter. Cool the filtrate, and add 2 to 3 drops of gelatin TS: a white turbidity or precipitate is produced.

(2) Shake 0.1 g of pulverized Gambir with 20 mL of dilute ethanol for 2 minutes, and filter. Mix 1 mL of the filtrate with 9 mL of dilute ethanol, and to the solution add 1 mL of vanillin-hydrochloric acid TS: a light red to red-brown color develops.

Total ash Not more than 6.0%.

Acid-insoluble ash Not more than 1.5%.

**Extract content** Dilute ethanol-soluble extract: not less than 70.0%.

#### **Powdered Gambir**

Gambir Pulveratum

アセンヤク末

Powdered Gambir is the powder of Gambir.

**Description** Powdered Gambir occurs as a red-brown to dark brown powder. It has a slight odor, and an extremely astringent and bitter taste.

Under a microscope, Powdered Gambir, immersed in olive oil or liquid paraffin, consists of needle crystalline masses or yellow-brown to red-brown angular fragments, and reveals epidermal tissue and thick-walled hairs.

Identification (1) To 0.2 g of Powdered Gambir add 10

mL of water, warm in a water bath for 5 minutes with occasional shaking, and filter. Cool the filtrate, and add 2 to 3 drops of gelatin TS: a white turbidity or precipitate is produced.

(2) Shake 0.1 g of Powdered Gambir with 20 mL of dilute ethanol for 2 minutes, and filter. Mix 1 mL of the filtrate with 9 mL of dilute ethanol, and to the solution add 1 mL of vanillin-hydrochloric acid TS: a light red to red-brown color develops.

Total ash Not more than 6.0%.

Acid-insoluble ash Not more than 1.5%.

**Extract content** Dilute ethanol-soluble extract: not less than 70.0%.

## $\beta$ -Galactosidase (Aspergillus)

#### Aspergillus Galactosidase

 $\beta$ -ガラクトシダーゼ(アスペルギルス)

[9031-11-2]

 $\beta$ -Galactosidase (Aspergillus) contains an enzyme produced by *Aspergillus oryzae*. It is an enzyme drug having lactose decomposition activity, and contains 8000 to 12000 units per g. Usually, it is diluted with a mixture of maltose and dextrin, maltose and D-mannitol, or maltose, dextrin and D-mannitol.

**Description**  $\beta$ -Galactosidase (Aspergillus) occurs as a white to light yellow powder.

It is slightly soluble in water with a turbidity, and practically insoluble in ethanol (95) and in diethyl ether.

**Identification** (1) Dissolve 0.025 g of  $\beta$ -Galactosidase (Aspergillus) in 100 mL of water, then to 1 mL of this solution add 9 mL of lactose substrate TS, and stand at 30°C for 10 minutes. To 1 mL of this solution add 6 mL of glucose detection TS, and stand at 30°C for 10 minutes: a red to redpurple color develops.

(2) Dissolve 0.1 g of  $\beta$ -Galactosidase (Asperigillus) in 100 mL of water, and filter the solution if necessary. Determine the absorption spectrum of the solution as directed under the Ultraviolet-visible Spectrophotometry, and compare the spectrum with the Reference Spectrum: both spectra exhibit similar intensities of absorption at the same wavelengths.

**Purity** (1) Odor— $\beta$ -Galactosidase (Aspergillus) has no any rancid odor.

- (2) Heavy metals—Proceed with 1.0 g of  $\beta$ -Galactosidase (Aspergillus) according to Method 2, and perform the test. Prepare the control solution with 2.0 mL of Standard Lead Solution (not more than 20 ppm).
- (3) Arsenic—Prepare the test solution with 1.0 g of  $\beta$ -Galactosidase (Aspergillus) according to Method 3, and perform the test using Apparatus B (not more than 2 ppm). **Loss on drying** Not more than 9.0% (0.5 g, in vacuum, 80°C, 4 hours).

**Residue on ignition** Not more than 3.0% (0.5 g).

Nitrogen content Weigh accurately about 0.07 g of  $\beta$ -Galactosidase (Aspergillus), and perform the test as directed under the Nitrogen Determination: the amount of nitrogen (N: 14.007) is between 0.5% and 5.0%, calculated on the dried basis

- Assay (i) Substrate solution: Dissolve 0.172 g of 2-nitrophenyl- $\beta$ -D-galactopyranoside in disodium hydrogen-phosphate-citric acid buffer solution, pH 4.5 to make 100 mL.
- (ii) Procedure: Weigh accurately about  $0.025 \, \mathrm{g}$  of  $\beta$ -Galactosidase (Aspergillus), dissolve in water to make exactly  $100 \, \mathrm{mL}$ , then pipet  $2 \, \mathrm{mL}$  of this solution, add water to make exactly  $50 \, \mathrm{mL}$ , and use this solution as the sample solution. Take exactly  $3.5 \, \mathrm{mL}$  of the substrate solution, stand at  $30 \pm 0.1 \, ^{\circ}\mathrm{C}$  for  $5 \, \mathrm{minutes}$ , add exactly  $0.5 \, \mathrm{mL}$  of the sample solution, immediately mix, and stand at  $30 \pm 0.1 \, ^{\circ}\mathrm{C}$  for exactly  $10 \, \mathrm{minutes}$ , then add exactly  $1 \, \mathrm{mL}$  of sodium carbonate TS and mix immediately. Determine the absorbance,  $A_1$ , of this solution at  $420 \, \mathrm{nm}$  using water as the control. Separately, take exactly  $3.5 \, \mathrm{mL}$  of the substrate solution, add exactly  $1 \, \mathrm{mL}$  of sodium carbonate TS and mix, then add exactly  $1 \, \mathrm{mL}$  of the sample solution and mix. Determine the absorbance,  $A_2$ , of this solution in the same manner as above.

Units per g of 
$$\beta$$
-Galactosidase (Aspergillus)  
=  $\frac{A_1 - A_2}{0.917} \times \frac{1}{0.5} \times \frac{1}{10} \times \frac{1}{W}$ 

0.917: Absorbance of 1 μmol/5 mL of o-nitrophenol
W: Amount (g) of the sample in the sample solution per mL

Unit: One unit indicates an amount of the enzyme which decomposes 1  $\mu$ mol of 2-nitrophenyl- $\beta$ -D-galactopyranoside in 1 minute under the above conditions.

**Containers and storage** Containers—Tight containers. Storage—In a cold place.

# **β-Galactosidase (Penicillium)**

β-ガラクトシダーゼ(ペニシリウム)

[9031-11-2]

β-Galactosidase (Penicillium) contains an enzyme, having lactose decomposition activity, produced by *Penicillium multicolor*. It contains not less than 8500 units and not more than 11,500 units in each g. Usually, it is diluted with D-mannitol.

**Description**  $\beta$ -Galactosidase (Penicillium) occurs as a white to pale yellowish white, crystalline powder or powder.

It is soluble in water with a turbidity, and practically insoluble in ethanol (95).

It is hygroscopic.

**Identification** (1) Dissolve 0.05 g of  $\beta$ -Galactosidase (Penicillium) in 100 mL of water, then to 0.2 mL of this solution add 0.2 mL of lactose substrate TS, and allow to stand at 30°C for 10 minutes. To this solution add 3 mL of glucose detection TS, and allow to stand at 30°C for 10 minutes: a red to red-purple color develops.